### Antidiabetic, hepato-protective and hypolipidemic effects of *Eryngium caucasicum* extract in streptozotocin-nicotinamide induced type 2 diabetes in male rats

Masood Afshari<sup>1</sup>, Majid Mohammadshahi<sup>2</sup>, Ali Reza Malayeri<sup>3\*</sup>, and Laleh Zaheri<sup>4</sup>

<sup>1</sup>Department of Nutrition, Jundishapour University of Medical Sciences, Ahvaz, Iran.

<sup>2</sup>Department of Nutrition, Hyperlipidemia Research Center, Jundishapour University of Medical Sciences. Ahvaz, Iran.

<sup>3</sup>Medical Plant Research Center, Jundishapour University of Medical Sciences. Ahvaz, Iran.

<sup>4</sup>Department of Persian Pharmacy Faculty of Pharmacy, Jundishapour University of Medical Sciences. Ahvaz, Iran.

Correspondence to Ali Reza Malayeri (email: shahi334@gmail.com).

(Submitted: 18 December 2018 – Revised version received: 12 January 2019 – Accepted: 29 January 2019 – Published online: 26 March 2019)

**Objective** The purpose of this study was to evaluate anti-diabetic, hepato-protective and hypolipidemic effects of *Eryngium caucasicum* extract in nicotinamide-streptozotocin induced type 2 diabetes mellitus model in Wistar rats.

**Methods** In this study, sixty adult male Wistar rats (150–250 g) were randomly allocated into six groups (n = 10) including: (1) healthy control rats, (2) diabetic control, (3) diabetic rats which received Sitagliptin, (4–6) diabetic rats which received respectively 100, 200 and 300 mg/kg oral gavage of *E. caucasicum* extract for 30 days. Diabetes was induced by intraperitoneal injection of nicotinamide and streptozotocin. At the end of experiment, blood samples were collected and fasting blood sugar, insulin, hepatic enzymes, lipid profile and insulin-like growth factor were measured.

**Results** Administration of *E. caucasicum* in type 2 diabetes mellitus animal models can affect insulin secretion, insulin related indices and lipid profile. Liver enzymes not only did not increase but also decreased significantly. Additionally, *E. caucasicum* increased insulin-like growth factor serum level in a dose-dependent manner. On the whole, it seems that *E. caucasicum* affects diabetes in rats in a dose-dependent manner, as higher doses showed better effects.

**Conclusion** Here, we showed that administration of *E. caucasicum* in type 2 diabetes mellitus animal model has antidiabetic, hypolipidemic and hepato-protective effects which may be in a dose-dependent manner.

Keywords E. caucasicum extract, insulin, lipid profile, liver enzymes, type 2 diabetes mellitus, antidiabetic, hepato-protective, hypolipidemic

### Introduction

Diabetes mellitus (DM) is a multifactorial chronic disorder characterized by persistent hyperglycemia in which insulin becomes disabled to perform main function due to weakened insulin secretion, insulin resistance or both.1 DM is recognized as one of the most prevalent endocrine disorders which have reached epidemic proportions around the world. Based on WHO reports in 2030 diabetes patients will be 333 million or 6.3% of the total population in the world.<sup>2,3</sup> Type 2 DM (T2DM) is more prevalent.<sup>4</sup> DM devoted a major challenge to healthcare systems around the world with about 7–13% of the healthcare budget of the worldwide.<sup>5,6</sup> Prolonged hyperglycemia in DM leads to many chronic complications such as vascular diseases, retinopathy, neuropathy and nephropathy which increase mortality and morbidity among patients with DM.7 DM related abnormal lipid metabolism is considered as a major reason for development of atherosclerosis and cardiovascular complications.8

Growing evidence suggests that inflammation involved in T2DM pathogenesis, insulin resistance and diabetes-related vascular complications.<sup>9</sup> Hyperglycemia and increased free fatty acids (FFA) may promote inflammation through inducing glucose utilization which accompanied by changes in oxidative phosphorylation.<sup>10,11</sup> This setting induces proinflammatory trait in macrophages invading the adipose tissue and other tissues including the pancreatic islets and vascular cells.<sup>12</sup> Glucotoxicity and lipotoxicity also exert oxidative stress and increase the release of active interleukin (IL-1 $\beta$ ).<sup>10,11</sup> IL-1 $\beta$  *in situ* intensifies inflammation through expression of

different cytokines in diabetic  $\beta$ -cells, adipose tissues and blood vessels.<sup>13</sup> So, it has been postulated that targeting inflammation not only attenuates DM but also could prevent its progression and vascular complications.<sup>14</sup>

Despite several drugs against diabetes, due to unpleasant side effects like peripheral edema, gastrointestinal discomfort and hypoglycemia, the need for newer drugs with fewer side effects raised.<sup>15</sup> Presently, there is mounting interest in herbal medicines for the treatment of T2DM because of their perceived effectiveness, higher safety and relatively low costs.<sup>16</sup> It has been estimated that more than 1000 herbal remedies are being used against DM.<sup>17</sup> The chemical composition of plants have been implicated as their biological effects to treat DM. Compounds like flavonoids, terpenoids, phenolic compounds and other constituents which show fasting blood glucose (FBS) lowering effects.<sup>18,19</sup>

The *Eryngium* genus described as the most taxonomically complex and the largest genus of family of Apiaceae (Umbelliferae). *Eryngium* L. consists of more than 250 species in Eurasia, North Africa, America and Australia.<sup>20,21</sup> *Eryngium caucasicum* which is known as Eryngo is cultivated in middle-east countries like Iran and Turkey.<sup>22</sup> In northern Iran this extract has been widely used as different foodstuffs like pickles.<sup>23</sup> Several therapeutic uses were reported for Apiaceae such as stimulant, diaphoretic, diuretic, stone inhibitor, expectorant and anthelmintic.<sup>24</sup> In Turkish folk medicine, it has been used as a remedy against various inflammatory disorders, edema, sinusitis, urinary infections or inflammations etc.<sup>21</sup> *Eryngium* 

*caucasicum* has been used as a reinforcing generative power, diuretic, lenitive and appetizer.<sup>25,26</sup> Antioxidant, antihypoxic and free radical scavenging and reno-protective effects of *Eryngium caucasicum* Trautv. leaves have been reported.<sup>23,27,28</sup> Antiinflammatory activities of other *Eryngium* species cultivated in Turkey have been assessed *in vivo*. Aerial parts and roots reported to possess most activities.<sup>29</sup> Several bioactive compounds, mainly phenolic compounds and terpenoids have been purified from *Eryngium* species which are comprising triterpenoid saponins, flavonoids, coumarins and acetylenes.<sup>30</sup>

In this study, we aimed to investigate antidiabetic, hypolipidemic and hepato-protective potentials of Eryngo on nicotinamide-streptozotocin (NA-STZ)-induced T2DM model in rats.

### **Materials and Methods**

## Preparation of E. caucasicum leaves hydro-alcoholic extract

In this experimental study, fresh leaves of E. caucasicum were collected from Mazandaran province, Iran. Department of the Botany of Ahvaz Jundishapur University of Medical Science (AJUMS) scientifically validated the plant. Voucher specimens are deposited in the Herbarium Department of Biology, University of Mazandaran, Iran (No. 1442). The leaves were dried in shade and afterward were crushed and milled by electrical grinders. To prepare E. caucasicum leaves of hydro-alcoholic extract, 300 g of E. caucasicum leaves powder was dissolved in 1200 mL of a distilled water:ethanol mixture (70:30) and were stored for 72 h at room temperature. After filtering the mixture with Whatman No. 1 filter paper, the mixture was centrifuged at 3500 rpm for 20 min. Condensation was performed by rotary evaporator. Eventually, the supernatant was removed and dried at 37°C, and the obtained semisolid mass was stocked in refrigerator until injections.

### Animals

Sixty adult male Wistar rats between 150 and 250 g were obtained from animal house of AJUMS. All animals were kept in temperature about  $22 \pm 2^{\circ}$ C and 12 h light/12 h dark cycle. All animals were permitted *ad libitum* access to normal laboratory diet and tap water. Maintenance and care of experimental animals complied with the National Research Council of the National Academic.

### Induction of type 2 diabetes model

Induction of type 2 diabetes was first performed by an intraperitoneal (IP) injection of a single-dose of NA (120 mg/kg) (dissolved in normal saline) (Sigma-Aldrich Co., St Louis, MO, USA) dissolved in distilled water. After 15 min, a single IP dose of STZ (55 mg/kg) (dissolved in citrate buffer, pH 4.5, 0.1M) (Sigma-Aldrich Co., St Louis, MO, USA) were also injected. The rats were fasted overnight before the induction. The development of T2DM was evaluated by assaying FBS 72 h after NA–STZ injection. FBS higher than 250 mg/dL was considered as diabetes. The other animals with lower serum glucose levels were removed from the experiments.

### Study design

All the rats were divided into six groups with 10 rats in each group as follows:

Group I:	10 healthy control rats (0.5 mL/kg normal					
	saline oral gavage).					
Group II:	10 diabetic rats without treatment (0.5 mL/					
	kg normal saline oral gavage).					
Group III:	10 diabetic rats treated with Sitagliptine					
-	(10 mg/kg oral gavage) as a standard drug					
	for diabetes.					
Groups IV–VI:	10 diabetic rats that received hydro-					
	alcoholic extract of E. caucasicum at 100,					
	200 and 300 mg/kg oral gavage respec-					
	tively. Body weight was recorded weekly.					

Sitagliptine and the hydro-alcoholic extract of *E. caucasicum* were administered for 30 days. Sitagliptine was dissolved in distilled water. About 20 h before the end of experiment, animals were deprived of food overnight. All the rats were anesthetized by ether. Blood samples were collected by cardiac puncture. To separate serum, blood samples were centrifuged at 3500 rpm for 20 min. Then, serum samples were stored at  $-80^{\circ}$ C until biochemical measurements.

#### **Biochemical measurements**

Fasting blood glucose was measured by Elegance glucometer (CT-X10, Convergent Technologies, Germany) through the lateral tail vein of rats in days 0 and 28. Circulating insulin level was evaluated ELISA kit (Mercodia Corporation, Uppsala, Sweden). The kit has assay sensitivity of 1 mIU/mL. Intra- and inter-assay coefficient of variation (CV) were of 2.1% and 6.5% respectively.

Also, insulin resistance (homeostasis model assessment of insulin resistance; HOMA-IR) and quantitative insulin sensitivity check index (QUICKI) were calculated by the following formula:

	Fasting blood glucose (mg/dL) $\times$			
HOMA-IR:	Insulin (µIU/mL)			
	405			
QUICKI: (1	1			
	og FBS (mg/dL) + log insulin (μIU/mL))			

The serum level of TC, low-density lipoprotein (LDL), triglyceride (TG), high-density lipoprotein (HDL), enzyme activity of alkaline phosphatase (ALP), aspartate aminotrans-ferase (AST) were assessed by auto-analyzer using commercial kits (Pars Azmoon, Iran). CV for all kits were lower than 7% for intra- and inter-assay.

Insulin-like growth factor (IGF-1) serum level also was assessed by chemiluminescence immunoassay method (Diasorin, Italy). CV for intra- and inter-assay were <0.05.

### Statistical analysis

All data were expressed as mean  $\pm$  standard error of mean (SEM). Differences between all groups were made by one-way analysis of variance (ANOVA) followed by post-hoc test [least significant difference (LSD)]. Statistically significant was considered at *P* < 0.05. All Statistical analysis was performed by SPSS Statistics V. 17.01 (SPSS Inc., Chicago, USA).

### Results

# Effects of hydro-alcoholic extract E. caucasicum on body weight

As depicted in Table 1, STZ–NA diabetes induction, did not change body weight in diabetic control group compared with healthy control group (P = 0.634). However, administration of Sitagliptine in diabetic rats lead to significant reduction of body weight in comparison with diabetic control and healthy control group (P = 0.008 and 0.028 respectively). After administration of *E. caucasicum* extract in diabetic rats (at dose of 300 mg/kg) body weight increased significantly in comparison with Sitagliptin group (P = 0.002).

# Effect of hydro-alcoholic extract E. caucasicum on fasting blood glucose, insulin level, and insulin-related indexes

Induction of type 2 diabetes using STZ–NA significantly increased FBS in diabetic rats compared with healthy control group (P = 0.009). Sitagliptin also decreased FBS in diabetic rats significantly (P = 0.004). Furthermore, daily administration of hydro-alcoholic *E. caucasicum* in diabetic rats revealed a dose-dependent significant reduction in FBS in distinct period. *Eryngium caucasicum* administration at 200 and 300 mg/kg doses cause significant reduction in FBS compared with diabetic control group (P = 0.038 and 0.004 respectively). In essence although *E. caucasicum* administration at dose of 100 mg/kg revealed no significant change, but reducing trend reported following increasing doses of *E. caucasicum* extract as shown in Table 1.

Induction of diabetes in Wistar rats lead to remarkable decrease in insulin serum level compared with control group (P < 0.001). Administration of Sitagliptin reversed reduction in insulin significantly (P = 0.008). In addition three different doses of *E. caucasicum* extract also improved insulin serum level in diabetic rats (*P*-value for all doses <0.001). HOMA-IR the index of insulin resistance were lower in three different doses of *E. caucasicum* extract compared with healthy control group at the end of intervention (*P*-value <0.01 for all doses). Sitagliptin administration increased HOMA-IR index significantly compared with diabetic control group (P = 0.006). Also there were significant difference in dose of 300 mg/kg and Sitagliptin at the end of trial (P = 0.011). QUICKI, other index which shows the health of insulin-producing cells, revealed significant increase in diabetic rats compared with control

group (P < 0.001). This increase in QUICKI also was significant for 300 mg/kg dose of *E. caucasicum* compared with control and Sitagliptin group. Only Sitagliptin group 100 and 200 mg/kg dose of *E. caucasicum* showed significant reduction compared with diabetic control group (P = 0.002, 0.047 and 0.014 significantly).

## Effect of hydro-alcoholic extract E. caucasicum on lipid profile and liver enzymes

As demonstrated in Table 2, regarding HDL marginal significant difference were seen only in the 200 mg/kg dose of *E. caucasicum* extract and Sitagliptin group (P = 0.051). There were no significant differences between other experimental groups. LDL cholesterol decreased significantly in the Sitagliptin and 300 mg/kg dose of E. caucasicum compared with diabetic control group (P-value <0.05 in both groups). It must be noted that induction of diabetes in Wistar rats increased TC in diabetic control group compared with healthy control group (P = 0.020). Treatment of diabetic rats with Sitagliptin and E. caucasicum extract in doses of 100 and 300 mg/kg body weight decreased TC significantly compared with diabetic control group (P < 0.01). Administration of Sitagliptin and E. caucasicum extract in doses of 200 and 300 mg/kg improved serum TG compared with diabetic control group (P < 0.05). Also as depicted in Table 2, dose of 300 mg/kg decreased TG serum level more than Sitagliptine group at the end of trial (P = 0.003).

Results showed that after induction of diabetes by STZ– NA, serum alanine aminotransferase (ALT) and AST levels were increased compared with control group (P = 0.020). Although Sitagliptin and all three experimental doses of plant improved ALT in diabetic rats, but regarding AST dose of 300 mg/kg decreases serum level of AST (P = 0.005).

### Effect of hydro-alcoholic extract E. caucasicum on IGF-1 serum level

As depicted in Fig. 1, induction of diabetes resulted in significant decrease in IGF-1 in all experimental diabetic rats compared with control group (P < 0.01). Treatment with Sitagliptine improved serum level of IGF-1 compared with diabetic control group. Administration of plant extract at 100 and 200 mg/kg doses were not efficient as Sitagliptin treatment for improving IGF-1 serum level. However, again in a dose-dependent manner 300 mg/kg of herb extract increased serum level of IGF-1 significantly (P < 0.001) which was also higher than first dose (P = 0.026).

Table 1. Effect of hydro-alcoholic extract of *E. caucasicm* (E.C.) on body weight, fasting blood glucose, insulin, HOMA-IR and QUICKI in nicotinamide–streptozotocin-induced diabetic Wistar rats

	Healthy control	Diabetic control	Diabetes ± Sitagliptin	Diabetes ± 100 mg/kg E.C.	Diabetes ± 200 mg/kg E.C.	Diabetes ± 300 mg/kg E.C.
Weight (g)	221.08 ± 6.96	$216.52 \pm 7.04$	$193.67 \pm 6.45^{a,b}$	$240.28\pm6.87^{\circ}$	$209.30 \pm 6.55$	211.12 ± 6.35
FBS (mg/dL)	107.50 ± 34.5	157.28 ± 59.00ª	$104.57 \pm 22.32^{\rm b}$	130.16 ± 18.49	120.42 ± 16.25 <sup>b</sup>	103.85 ± 17.72 <sup>b</sup>
Insulin (µU/mL)	$7.04 \pm 0.99$	$2.16\pm0.66^{\text{a}}$	$5.75\pm0.82^{\text{a,b}}$	$3.46\pm0.61^{a,b,c}$	$4.17\pm0.92^{\text{a,b,c}}$	$3.54\pm0.86^{\text{a,b,c}}$
HOMA-IR	$0.190\pm0.080$	$0.084\pm0.038^{\text{a},}$	$0.150 \pm 0.041^{\rm b}$	$0.11\pm0.024^{\text{a}}$	$0.122\pm0.023^{\text{a}}$	$0.089 \pm 0.019^{a,c}$
QUICKI	$0.352 \pm 0.021$	$0.406 \pm 0.041^{a}$	$0.362 \pm 0.021^{\rm b}$	$0.378 \pm 0.012^{\rm b}$	$0.372 \pm 0.013^{\rm b}$	$0.393 \pm 0.019^{a,c}$

 $^{\circ}P < 0.05$  compared with the control group.  $^{\circ}P < 0.05$  compared with the diabetic control group.  $^{\circ}P < 0.05$  compared with the diabetic animals received Sitagliptin. E.C., Eryngium caucasicm; FBS, fasting blood glucose; HOMA-IR, the homeostatic model assessment-insulin resistance; QUICKI, quantitative insulin-sensitivity check index. Results are expressed as mean  $\pm$  standard error of mean (SEM), one-way ANOVA and post-hoc LSD test.

· ·						
	Healthy control	Diabetic control	Diabetes ± Sitagliptin	Diabetes ± 100 mg/kg E.C.	Diabetes ± 200 mg/kg E.C.	Diabetes ± 300 mg/kg E.C.
LDL (mg/dL)	20.83 ± 7.16	29.91 ± 19.22	$17.97 \pm 4.70^{ m b}$	21.73 ± 4.12	23.80 ± 6.82	$18.11 \pm 4.95^{\text{b}}$
HDL (mg/dL)	$50.30 \pm 4.23$	$33.71 \pm 5.12$	$30.14 \pm 4.81$	$30.66 \pm 3.82$	$35.14 \pm 6.36^{\circ}$	$31.28 \pm 2.05$
TC (mg/dL)	$58.83 \pm 10.88$	$72.85 \pm 16.49^{\circ}$	$54.28\pm9.08^{\rm b}$	$58.83 \pm 5.77^{\rm b}$	$65.00 \pm 10.63$	$54.28 \pm 3.35^{\rm b}$
TG (mg/dL)	$37.50 \pm 9.43$	46.14 ± 24.16	$30.85 \pm 9.63^{\rm b}$	32.16 ± 8.20	$30.28\pm8.61^{\rm b}$	$24.42\pm8.24^{\rm b}$
ALT (mg/dL)	$45.83 \pm 7.46$	$77.28 \pm 12.86^{\circ}$	57.28 ± 17.13 <sup>b</sup>	$52.16 \pm 26.79^{\text{b}}$	$44.57 \pm 11.64^{\text{b}}$	47.14 ± 17.41 <sup>b</sup>
AST (mg/dL)	115.50 ± 14.54	156.14 ± 24.65ª	138.71 ± 20.28	141.50 ± 48.09	137.42 ± 35.42	$109.00 \pm 24.93^{ m b}$

Table 2. Effect of hydro-alcoholic extract of *E. caucasicm* (E.C.) on lipid profile and liver enzymes in nicotinamidestreptozotocin-induced diabetic Wistar rats

 $^{a}P < 0.05$  compared with the control group.  $^{b}P < 0.05$  compared with the diabetic control group.  $^{C}P < 0.05$  compared with the diabetic animals received Sitagliptin. Results are expressed as mean  $\pm$  standard error of mean (SEM), one-way ANOVA and post-hoc LSD test. E.C., *E. caucasicm*; ALT, alanine aminotransferase; AST, aspartate aminotransferase; LDL, low-density lipoprotein; HDL, high-density lipoprotein; TC, total cholesterol; TG, triglyceride.

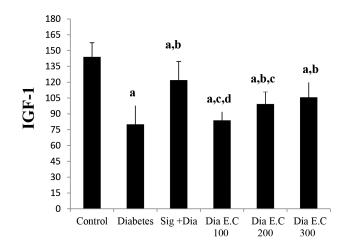


Fig. 1 Effect of hydro-alcoholic extract of *E. caucasicm* (E.C.) on serum insulin-like growth factor (IGF-1) level in nicotinamide– streptozotocin-induced diabetic Wistar rats. Values outlined as mean  $\pm$  standard error of mean (SEM), One-way ANOVA and posthoc LSD test. Dia, diabetic; E.C., *E. caucasicm*; Sig, Sitagliptin; 100, 100 mg/kg hydro-alcoholic extract E.C.; 200, 200 mg/kg hydro-alcoholic extract E.C.;  $^{a}P < 0.05$  compared with the control group.  $^{b}P < 0.05$  compared with the diabetic animals received Sitagliptin.  $^{d}P < 0.05$  compared with the diabetic animals received 300 mg/kg hydro-alcoholic extract E.C.

### Discussion

In this study, we investigated the effect of *E. caucasicum* in STZ–NA rats, T2DM animal model. In this study, consistent with reports of earlier investigators insulin serum level reduced significantly in diabetic group. STZ increase reactive oxygen species generation through intracellular nitric oxide (NO) and lead to  $\beta$ -cell dysfunction.<sup>31</sup> Our result showed that *E. caucasicum* extract significantly decrease FBS in high doses (200 and 300 mg/kg of Eryngo extract). Administration of *E. caucasicum* in diabetic rats also improved insulin secretion significantly. The insulin secretary effects of *E. caucasicum* were higher than Sitagliptin as well. The possible mechanism related to phenolic content of *E. caucasicum* extract to restore the function of pancreatic tissue. Anti-diabetic potential of phenolic compounds has been widely investigated.<sup>32</sup>

control through multiple mechanisms.33,34 Enhanced insulin-mediated glucose uptake through glucose transporter 4-mediated process,<sup>35-37</sup> improvement of pancreatic  $\beta$ -cell function through antioxidant activity and liver glucose homeostasis.<sup>38,39</sup> In addition inflammation is among several pathways which have been suggested for anti-diabetic effects of phenolic compounds.33 One of interesting mechanisms is through the potential prebiotic effects of dietary polyphenols to increase the population of Bifidobacteria which has been associated with improved glucose tolerance and lowered inflammatory biomarkers such as the IL-6, and IL-1 $\beta$  and tumor necrosis factor a (TNF-a).40,41 Previous studies reported that Eryngium species are rich medicinal plant of phenolic and flavonoids compounds with antioxidant and anti-inflammatory activities.<sup>42-47</sup> Also, saponins have blood glucose lowering effect which also found especially in E. caucasicum.48 The insulin-related indexes including QUICKI and HOMA-IR also were calculated. HOMA-IR reported as a strong clinical and epidemiological tool for assessment of insulin resistance.48 Hence, these results indicate that E. caucasicum extract have favorable effects on QUICKI compared with diabetic group; however despite the insulin secretary effect of E. caucasicum, HOMA-IR not showed favorable changes after administration of E. caucasicum extract. On the other hand several studies have evaluated the effect of IGF-I on insulin sensitivity and T2DM.49 Large longitudinal studies reported a higher risk of insulin resistance, metabolic syndrome and worse glycemic control in subjects with low IGF-I serum concentrations.<sup>50-53</sup> Here, we showed that administration of E. caucasicum dose-dependently increase circulating IGF-1 in diabetic rats. The biological mechanism and possible effect has to be elucidated in more detail in future studies.

The liver is a vital insulin-dependent organ and is severely affected during diabetes mellitus.<sup>54,55</sup> Hepatocellular damage in T2DM model described by raise in serum level of ALT and AST.<sup>56</sup> In our investigation the serum level of liver enzymes also increased significantly in diabetic group compared with control group. The reversal of ALT and AST activity toward near normalcy interpreted as the prevention of cellular and tissue damage under diabetic conditions.<sup>55,57,58</sup> The *E. caucasicum* extract caused a significant reduction in level of AST and ALT in treated diabetic groups which shows its hepato-protective effects. ALT improved in all doses and AST only in highest dose. Previously reno-protective effects of *E. caucasicum* extract has been contributed to its ability to scavenge free radical.<sup>59</sup> Other species of Eryngium family also showed hepato-protective effects. Especially one study demonstrated that *E. caucasicum* Trautv. extract due to the presence of high phenolic compounds has protective effects on tricyclazole induced hepatotoxicity.<sup>60</sup> The antioxidant effect of *E. caucasicum* extract is suggested.

Reduced lipoprotein lipase activity lead to unbalanced lipoprotein metabolism during diabetes.<sup>61,62</sup> On the other hand free fatty acid influx stimulates hepatic triglyceride synthesis and over production of LDL.63 Also a noticeable increase in sterol regulatory element-binding protein-1 and fatty acid synthase expression in STZ-diabetic rats has been reported.<sup>64</sup> LDL and TG serum level improved only in highest dose of E. caucasicum. HDL did not show any significant effect however in comparison with Sitagliptin, 200 mg/kg of E. caucasicum lead to higher HDL serum level. TC also improved in lowest and highest doses. The probable mechanisms regarding dyslipidemia comprising stimulation of cholesterol-7-alpha-hydroxylase, which converts cholesterol into bile acids; inhibition of HMG-CoA reducates; inhibition of cholesterol absorption from the intestines due to the formation of complexes with compounds such as glycosides and saponins.<sup>65,66</sup> In addition increase in insulin secretion and changes lipolysis which are under the control of insulin.67 Saponins reported to stimulate the decline and excretion of plasma cholesterol, and improved lipid metabolism.68,69

### Previous study also reported that other Eryngium species improve lipid profile in diabetic rats.<sup>70</sup>

In recent years oxidative stress may represent the key role in the pathogenesis of secondary diabetic complication. Hyperglycemia is a main cause for elevated free radical levels and oxidative damage.<sup>71,72</sup> On the whole antioxidant and antiinflammatory effects of *E. caucasicum* could be contributed to possible protective effects against diabetes complication.

### Conclusion

In conclusion we showed that administration of *E. caucasicum* in T2DM animal model improved insulin secretion, insulin related indexes and lipid profile. Liver enzymes not only have not increased but also decreased significantly; IGF-1 serum level increased in dose dependent manner after administration of *E. caucasicum*. On the whole it seems that *E. caucasicum* impact on diabetes rats in a dose-dependent manner, as higher doses showed better effects. However in future studies other dose-escalating intervention must be performed; in which higher doses evaluated in order to clarify the exact dose-response effect of *E. caucasicum*. Also toxicity in diabetes must be elucidated in future studies.

### **Conflicts of Interest**

None.

#### References

- 1. Blair M. Diabetes mellitus review. Urol Nurs. 2016;36:27–36.
- 2. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care.
- 2004;27:1047–53.
   Rathmann W, Giani G. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care. 2004;27:2568–2569.
- Centers for Disease Control and Prevention. National diabetes statistics report, 2017. Centers for Disease Control and Prevention, Atlanta, GA, 2017.
- 5. International Diabetes Federation ID. *Diabetes Atlas*. 3<sup>ed</sup> Ed.; International Diabetes Federation, Brussels, Belgium, 2006.
- Zhang P, Zhang X, Brown J, Vistisen D, Sicree R, Shaw J, et al. Global healthcare expenditure on diabetes for 2010 and 2030. Diabetes Res Clin Pract. 2010;87:293–301.
- 7. Papatheodorou K, Papanas N, Banach M, Papazoglou D, Edmonds M. Complications of diabetes 2016. J Diabetes Res. 2016;2016:6989453.
- 8. Rivellese A, Vaccaro O, Patti L. The pathophysiology of lipid metabolism and diabetes. Int J Clin Pract. 2004;58:32–35.
- Akash MS, Rehman K, Chen S. Role of inflammatory mechanisms in pathogenesis of type 2 diabetes mellitus. J Cell Biochem. 2013;114:525–531.
- Böni-Schnetzler M, Boller S, Debray S, Bouzakri K, Meier DT, Prazak R, et al. Free fatty acids induce a proinflammatory response in islets via the abundantly expressed interleukin-1 receptor I. Endocrinology. 2009;150:5218–5229.
- Maedler K, Sergeev P, Ris F, Oberholzer J, Joller-Jemelka HI, Spinas GA, et al. Glucose-induced beta cell production of IL-1beta contributes to glucotoxicity in human pancreatic islets. J Clin Invest. 2002;110:851–860.
- 12. Richardson SJ, Willcox A, Bone AJ, Foulis AK, Morgan NG. Islet-associated macrophages in type 2 diabetes. Diabetologia. 2009;52:1686–1688.
- Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annu Rev Immunol. 2009;27:519–550.
- Pollack RM, Donath MY, LeRoith D, Leibowitz G. Anti-inflammatory agents in the treatment of diabetes and its vascular complications. Diabetes Care. 2016;39:S244–S252.
- Ghorbani A. Best herbs for managing diabetes: a review of clinical studies. Funding Brazilian J Pharma Sci. 2013;49:413–422.
- 16. Rao MU, Sreenivasulu M, Chengaiah B, Reddy KJ, Chetty CM. Herbal medicines for diabetes mellitus: a review. Int J PharmTech Res. 2010;2:1883–1892.

- Marles RJ, Farnsworth NR. Antidiabetic plants and their active constituents. Phytomedicine. 1995;2:137–189.
- Jung M, Park M, Lee HC, Kang YH, Kang ES, Kim SK. Antidiabetic agents from medicinal plants. Curr Med Chem. 2006;13:1203–1218.
- Ji HF, Li XJ, Zhang HY. Natural products and drug discovery: can thousands of years of ancient medical knowledge lead us to new and powerful drug combinations in the fight against cancer and dementia? EMBO Rep. 2009;10:194–200.
- 20. Calviño CI, Martínez SG, Downie SR. The evolutionary history of Eryngium (Apiaceae, Saniculoideae): rapid radiations, long distance dispersals, and hybridizations. Mol Phylogenet Evol. 2008;46:1129–1150.
- Wörz A. On the distribution and relationships of the South-West Asian species of *Eryngium* L.(*Apiaceae-Saniculoideae*). Turk. J. Bot. 2004; 28:85–92.
- 22. Nabavi S, Nabavi S, Alinezhad H, Zare M, Azimi R. Biological activities of flavonoid-rich fraction of *Eryngium caucasicum* Trautv. Eur Rev Med Pharmacol Sci. 2012;16:81–87.
- Eslami SH, Ebrahimzadeh MA, Hajizadeh Moghaddam A, Nabavi SF, Jafari N, Nabavi SM. Renoprotective effect of *Eryngium caucasicum* in gentamicininduced nephrotoxic mice. Arch Biol Sci. 2011;63:157–160.
- Cardozo E, Rubio M, Rojas LB, Usubillaga A. Composition of the essential oil from the leaves of *Eryngium foetidum* L. from the Venezuelan Andes. J Essen Oil Res. 2004;16:33–34.
- 25. Trautv P. Plants for a future, edible, medicinal and useful plants for a healthier world, *Eryngium caucasicum*. 2004.
- Semnani K, Azadbakht M, Houshmand A. Composition of the essential oils of aerial parts of *Eryngium bungei* Boiss. and *Eryngium caeruleum* M. B. Pharm Sci. 2003;43–48.
- 27. Motallebi Riekandeh S, Mazandarani M, Ebrahimzadeh MA, Zargari M. Antioxidant activities of *Eryngium caucasicum* inflorescence. Eur Rev Med Pharmacol Sci. 2016;20:946–949.
- Khalili M, Dehdar T, Hamedi F, Ebrahimzadeh MA, Karami M. Antihypoxic activities of *Eryngium caucasicum* and *Urtica dioica*. Eur Rev Med Pharmacol Sci. 2015;19:3282–3285.
- Küpeli E, Kartal M, Aslan S, Yesilada E. Comparative evaluation of the antiinflammatory and antinociceptive activity of Turkish Eryngium species. J Ethnopharmacol. 2006;107:32–37.

- Wang P, Su Z, Yuan W, Deng G, Li S. Phytochemical constituents and pharmacological activities of Eryngium L.(Apiaceae). Pharmaceutical Crops. 2012;3:99–120.
- 31. Sheweita SA, Mashaly S, Newairy AA, Abdou HM, Eweda SM. Changes in oxidative stress and antioxidant enzyme activities in streptozotocin-induced diabetes mellitus in rats: role of *Alhagi maurorum* extracts. Oxidative medicine and cellular longevity. 2016;2016:8.
- 32. Lin D, Xiao M, Zhao J, Li Z, Xing B, Li X, et al. An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. Molecules. 2016;21. pii: E1374.
- Kim Y, Keogh JB, Clifton PM. Polyphenols and glycemic control. Nutrients. 2016;8. pii: E17.
- Ali Asgar Md. Anti-diabetic potential of phenolic compounds: a review. Int J Food Prop. 2013;16:91–103.
- Ueda-Wakagi M, Mukai R, Fuse N, Mizushina Y, Ashida H. 3-O-acylepicatechins increase glucose uptake activity and GLUT4 translocation through activation of PI3K signaling in skeletal muscle cells. Int J Mol Sci. 2015;16:16288–16299.
- Pinent M, Blay M, Bladé MC, Salvadó MJ, Arola L, Ardévol A. Grape seedderived procyanidins have an antihyperglycemic effect in streptozotocininduced diabetic rats and insulinomimetic activity in insulin-sensitive cell lines. Endocrinology. 2004;145:4985–4990.
- Kurimoto Y, Shibayama Y, Inoue S, Soga M, Takikawa M, Ito C, et al. Black soybean seed coat extract ameliorates hyperglycemia and insulin sensitivity via the activation of AMP-activated protein kinase in diabetic mice. J Agric Food Chem. 2013;61:5558–5564.
- Waltner-Law ME, Wang XL, Law BK, Hall RK, Nawano M, Granner DK. Epigallocatechin gallate, a constituent of green tea, represses hepatic glucose production. J Biol Chem. 2002;277:34933–34940.
- Wolfram S, Raederstorff D, Preller M, Wang Y, Teixeira SR, Riegger C, et al. Epigallocatechin gallate supplementation alleviates diabetes in rodents. J Nutr. 2006;136:2512–2518.
- Cani PD, Possemiers S, Van de Wiele T, Guiot Y, Everard A, Rottier O, et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. Gut. 2009;58:1091–1103.
- Cani PD, Neyrinck AM, Fava F, Knauf C, Burcelin RG, Tuohy KM, et al. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. Diabetologia. 2007;50:2374–2383.
- 42. Erdem SA, Nabavi SF, Orhan IE, Daglia M, Izadi M, Nabavi SM. Blessings in disguise: a review of phytochemical composition and antimicrobial activity of plants belonging to the genus *Eryngium*. Daru. 2015;23:53.
- Singh S, Singh DR, Banu S, Salim KM. Determination of bioactives and antioxidant activity in *Eryngium foetidum* L: a traditional culinary and medicinal herb. Proc Natl Acad Sci, India Sec B Biol Sci. 2013;83:453–460.
- Zhang Z, Li S, Ownby S, Wang P, Yuan W, Zhang W, et al. Phenolic compounds and rare polyhydroxylated triterpenoid saponins from *Eryngium yuccifolium*. Phytochemistry. 2008;69:2070–2080.
- Rjeibi I, Saad AB, Ncib S, Souid S. Phenolic composition and antioxidant properties of *Eryngium maritimum* (sea holly). J Coast Life Med. 2017;5: 212–215.
- Darriet F, Andreani S, Cian MD, Costa J, Muselli A. Chemical variability and antioxidant activity of *Eryngium maritimum* L. essential oils from Corsica and Sardinia. Flav Frag J. 2014;29:3–13.
- Nebija F, Stefkov G, Karapandzova M, Stafilov T, Panovska TK, Kulevanova S. Chemical characterization and antioxidant activity of *Eryngium campestre* L., Apiaceae from Kosovo. Macedon Pharm Bull. 2009;55:22–32.
- Patel DK, Prasad SK, Kumar R, Hemalatha S. An overview on antidiabetic medicinal plants having insulin mimetic property. Asian Pac J Trop Biomed. 2012;2:320–330.
- Friedrich N, Thuesen B, Jørgensen T, Juul A, Spielhagen C, Wallaschofksi H, et al. The association between IGF-I and insulin resistance: a general population study in Danish adults. Diabetes Care. 2012:35:768–773.
- 50. Sandhu MS, Heald AH, Gibson JM, Cruickshank JK, Dunger DB, Wareham NJ. Circulating concentrations of insulin-like growth factor-I and development

of glucose intolerance: a prospective observational study. Lancet. 2002;359:1740–1745.

- Sierra-Johnson J, Romero-Corral A, Somers VK, Lopez-Jimenez F, Mälarstig A, Brismar K, et al. IGF-I/IGFBP-3 ratio: a mechanistic insight into the metabolic syndrome. Clin Sci (Lond). 2009;116:507–512.
- 52. Aguirre GA, De Ita JR, de la Garza RG, Castilla-Cortazar I. Insulin-like growth factor-1 deficiency and metabolic syndrome. J Transl Med. 2016;14:3.
- 53. Færch L, Juul A, Pedersen-Bjergaard U, Thorsteinsson B. Association of IGF1 with glycemic control and occurrence of severe hypoglycemia in patients with type 1 diabetes mellitus. Endocr Connect. 2012;1:31–36.
- Mohamed J, Nazratun Nafizah AH, Zariyantey AH, Budin SB. Mechanisms of diabetes-induced liver damage: the role of oxidative stress and inflammation. Sultan Qaboos Univ Med J. 2016;16:e132–e141.
- 55. Rajasekaran S, Ravi K, Sivagnanam K, Subramanian S. Beneficial effects of aloe vera leaf gel extract on lipid profile status in rats with streptozotocin diabetes. Clin Exp Pharmacol Physiol. 2006;33:232–237.
- Kumar Bhateja P, Singh R. Antidiabetic activity of *Acacia tortilis* (Forsk.) Hayne ssp. raddiana polysaccharide on streptozotocin-nicotinamide induced diabetic rats. Biomed Res Int. 2014;2014:572013.
- Kazemi S, Asgary S, Moshtaghian J, Rafieian M, Adelnia A, Shamsi F. Liverprotective effects of hydroalcoholic extract of allium hirtifolium boiss. In rats with alloxan-induced diabetes mellitus. ARYA Atheroscler. 2010;6:11–15.
- Adeyemi DO, Ukwenya VO, Obuotor EM, Adewole SO. Anti-hepatotoxic activities of Hibiscus sabdariffa L. in animal model of streptozotocin diabetes-induced liver damage. BMC Complement Altern Med. 2014;14:277.
- Eslami SH, Ebrahimzadeh MA, Moghaddam AH, Nabavi SF, Jafari N, Nabavi SM. Renoprotective effect of Eryngium caucasicum in gentamicin-induced nephrotoxic mice. Arch Biol Sci Belgrade. 2011;63:157–160. doi:10.2298/ ABS1101157E
- Fattahi E, Jahromi VH. Protective effects of *Eryngium caucasicum* Trautv hydroalcholic extract on tricyclazole induced hepatotoxicity in mice. Majallah-i dānishqāh-i'ulūm-i pizishkī-i Arāk. 2016;19:67–78.
- 61. Tomkin GH, Owens D. Diabetes and dyslipidemia: characterizing lipoprotein metabolism. Diabetes Metab Syndr Obes. 2017;10:333–343.
- 62. Vergès B. Pathophysiology of diabetic dyslipidaemia: where are we? Diabetologia. 2015;58:886–899.
- 63. Punitha D, Thandavamoorthy A, Arumugasamy K, Suresh SN, Danya U, Udhayasankar MR. Anti-hyperlipidemic effect of ethanolic leaf extract of Gmelina arborea in streptozotocin induced male wistar albino rats. Int J Life Sci Pharma Res. 2012;2:46.
- Sun L, Halaihel N, Zhang W, Rogers T, Levi M. Role of sterol regulatory element-binding protein 1 in regulation of renal lipid metabolism and glomerulosclerosis in diabetes mellitus. J Biol Chem. 2002;277: 18919–18927.
- Abou El-Soud NH, Khalil MY, Hussein JS, Oraby FSH, Hussein Farrag A. Antidiabetic effects of fenugreek alkaliod extract in streptozotocin induced hyperglycemic rats. J Appl Sci Res. 2007;3:1073–1083.
- Saini S, Sharma S. Antidiabetic effect of *Helianthus annuus* L., seeds ethanolic extract in streptozotocinnicotinamide induced type 2 diabetes mellitus. Int J Pharm Pharm Sci. 2013;5:382–387.
- Cheng D, Liang B, Li Y. Antihyperglycemic effect of *Ginkgo biloba* extract in streptozotocin-induced diabetes in rats. Biomed Res Int. 2013;2013:162724.
- Lee SO, Simons AL, Murphy PA, Hendrich S. Soyasaponins lowered plasma cholesterol and increased fecal bile acids in female golden Syrian hamsters. Exp Biol Med (Maywood). 2005;230:472–478.
- 69. Milgate J, Roberts DCK. The nutritional & biological significance of saponins. Nutr Res. 1995;15:1223–1249.
- Zarei A, Ashtiyani SC, Hamidizadeh S, Rezaei A, Ramezani M, Aalizadeh A. The study of the effects hydro-alcoholic extract of *Eryngium billardieri* on lipid profiles levels and liver and renal functions tests in hypercholesterolemic rats. J Chem Pharma Res. 2015;7:200–206.
- Matough FA, Budin SB, Hamid ZA, Alwahaibi N, Mohamed J. The role of oxidative stress and antioxidants in diabetic complications. Sultan Qaboos Univ Med J. 2012;12:5–18.
- 72. Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res. 2010;107:1058–1070.

This work is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported License which allows users to read, copy, distribute and make derivative works for non-commercial purposes from the material, as long as the author of the original work is cited properly.