Evaluation of the role of serum uric acid in patients with multiple sclerosis; An observational case—control study

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Objectives: To evaluate the role of uric acid (UA) in patients with multiple sclerosis (MS) as an investigational marker in true relapse events.

Methods: 108 patients with relapsing–remitting and secondary progressive MS, compared to case–control of about 120 patients (manifested other neurological diseases, OND) who were included in this study which was conducted from March 2008 to July 2009 at Baghdad Teaching Hospital multiple sclerosis clinic. A study protocol sheet was done and filled from the patient's database in the MS clinic. **Results:** In the overall MS group, serum UA levels were lower than in controls, the difference did reach statistical significance (p = 0.01). Serum UA was found to be lower in patients during relapse than when they were in remission. The mean serum UA level from patients after 1 month of follow up shows an inverse correlation with MS type, age, and EDSS score, and positive correlation with gender and clinical activity, but none of these correlations reach statistical significance.

Conclusion: The question whether reduced serum UA level in MS is a primary deficit or an epiphenomenon remains open. Despite the fact that UA level was lower in clinically documented active patients, a general mean UA decrease is evident also in clinically and MRI inactive MS patients as compared to OND.

Keywords: Multiple sclerosis, uric acid, nitric oxide, peroxynitrite.

Introduction

Multiple sclerosis (MS) is a disease of myelin, the insulating cover around the nerves of the central nervous system (CNS: brain, optic nerves, and spinal cord) that becomes damaged in MS. MS most commonly begins in young adulthood and affects about twice as many women as men.¹ Uric acid (UA) is a diprotic acid with pKa1=5.4 and pKa2=10.3.²

There are now at least three independent mechanisms by which UA can interact with the immune system. The induction of the nitric oxide synthase isoform (NOS-2) in the CNS commonly associated with cells of the macrophage-monocyte lineage is a characteristic feature of experimental allergic encephalomyelitis (EAE). Moreover, production of the free radical nitric oxide (NO) in CNS tissue of mice has been correlated with the development of clinical signs of the disease.^{3,4} Peroxynitrite (ONOO2), a potent oxidant that is formed by the rapid combination of NO with superoxide (O22), can be formed in an inflammatory response⁵ and can cause a variety of toxic effects, including lipid peroxidation⁶ and tyrosine nitration.^{7,8} It has been shown that treatment with UA, a naturally occurring compound that selectively binds and inactivates peroxynitrite,9 inhibits the onset of clinical disease in an acute, aggressive form of mouse EAE, a result expected only if peroxynitrite is the more toxic molecule.¹⁰ It is proposed that soluble products generated from infiltrating immune cells and glial cells contribute to the damage to myelin and oligodendroglia in MS.^{11,12} When NO and superoxide are formed simultaneously, they may react to form the powerful oxidant peroxynitrite (ONOO-) at a rate constant which is three times faster than that at which superoxide dismutase scavenges superoxide.13 Extensive peroxynitrite activity has also been identified during early stages of EAE.14 UA is a strong peroxynitrite scavenger. The administration of UA to treat EAE in mice has been shown to produce a strong beneficial effect.

peroxynitrite inhibit components of the mitochondrial respiratory chain leading, if damage is severe enough, to a cellular energy deficiency state.¹⁵ It has been demonstrated that both NO and peroxynitrite may possibly have a role in the process of demyelination by inducing oligodendrocyte death¹⁶ and through damage of the myelin sheath by inducing lipid peroxidation.17 Moreover, NO donors have been shown to cause reversible conduction block in both normal and demyelinated axons of the central and peripheral nervous systems.^{18,19} Beneficial results of treatment with UA have been observed in experimental pneumococcal meningitis (PN)²⁰ and in focal brain injury in rats.²¹ By virtue of its potent oxidant activity, PN is believed to be responsible for the majority of damage to the CNS attributed to NO.22 Since observations on EAE may apply to MS one could expect that levels of UA, a natural scavenger of PN able to penetrate and preserve the blood-brain barrier integrity,²³ may be abnormal in MS patients. Regardless of clinical status, MS patients have significantly lower serum UA levels than controls.^{19,24} One study indicated that UA reduction is strictly linked to clinical and MRI activity in MS patients.²⁵ It is not known whether UA reduction is primarily linked to MS or simply represents an epiphenomenon.

It has been confirmed that NO and its toxic metabolite

Patients and methods

Blood samples were collected from 108 patients retrospectively diagnosed by revised McDonald criteria having definite MS. 94 patients from 108 patients had the relapsing–remitting (RR) type of the disease, 14 secondary progressives (SP). The patients selected randomly from the outpatient register of our Neurology Clinic Baghdad Teaching Hospital at Medical City for the period from March 2007 through July 2008.

The control population: we recruited 120 patients admitted to the neurological ward of Baghdad Teaching Hospital at Medical City the period from March 2007 through July 2008. There were 64 men and 56 women with other neurological diseases (ONDs) and clinical diagnosis (excluded gout). Our patients (cases and control) were sent for serum UA in fasting state. The cases with MS were followed up after one month to be another second reading of serum UA. Only 47 patients' serum UA results were documented in the protocol sheet. Serum UA levels of cases and controls were performed by the same laboratory of Baghdad Teaching Hospital, using a commercially available kit. Normal ranges of UA, according to the Baghdad Teaching Hospital at Medical City laboratory standardization, were 3.0–7.0 mg/dl.

Results

In the overall MS group, serum UA levels were lower than in controls, the difference did reach statistical significance (p = 0.01) as shown in Table 1.

There were 25 patients (from 108 patients) with clinically definite MS and their serum showed lower levels of UA (<3mg/ dl) compared to sera from controls with OND (4 patients from 120 patients) as shown in Table 1. 18 patients from 25 patients were RRMS while 7 patients were SPMS as shown in Table 2.

14 patients from 18 patients who had low serum UA were in clinically active disease (on relapse) and all patients with SPMS were in relapse (7 patients) as shown in Table 3.

14 patients from 18 patients with EDSS equal or more than 3.5 and all patients with SPMS with EDSS >3.5 as shown

in Table 4. In addition, in 47 RRMS patients in whom we performed repeated serum UA levels after 1-month analyses, we observed an increase in serum UA level during remission and decrease during relapse as shown in Table 5.

The mean serum UA level from patients after 1 month of follow up shows inverse correlation with MS type (-0.17), age of the patients (-0.239), and EDSS score (-0.21), and positive correlation with gender (0.11) and clinical activity (0.09); but none of these correlations reach statistical significance. We found a significant positive correlation of mean serum UA concentration before and after 1 month of follow up of those patients (p = 0.0001) as shown in Table 6.

Discussion

In the present study, we observed lower levels of UA in sera from patients with clinically active MS than in sera from clinically inactive MS patients or controls with OND. Additionally, in 47 RRMS patients in whom we performed repeated serum UA levels after 1-month analyses, we observed an increase in serum UA level during remission and decrease during relapse. We also found lower serum UA concentrations in the overall MS group than in the OND group, but the difference was statistically significant. UA levels in sera from MS patients have been recently reported in two studies.^{10,26} One found serum UA levels to be significantly lower in MS patients than in patients with OND,¹⁰ while the other observed no such difference.²⁶ However, the correlation between UA levels

Table 1. The percentage of low serum UA in the cases and control enrolled in the study.			
Number of patients	Number of patients with serum uric acid >3mg/dl	Number of patients with serum uric acid <3mg/dl	<i>p</i> -value
Cases (108)	83(77%)	25(23%)	0.01
Control (120)	116(97%)	4(3%)	0.05*
* N-+-::::			

* Not significant

Table 2. Relation between types of MS with the level of serum uric acid who their serum uric acid below 3 mg/dl.

MS type with number of patients	Number of patients with serum uric acid <3mg/dl	Number of patients with serum uric acid >3mg/dl
RRMS (94)	18 (19.1%)	76 (80.9%)
SPMS (14)	7 (50%)	7 (50%)
Total (108)	(25)	(83)

Table 3. The relation between types of MS who lowserum uric acid with active diseases (relapsing).

MS type with no. of patients	No. of patients in remission	No. of patients in relapsing	
RRMS 18	4 (22%)	14 (78%)	
SPMS 7	0 (0%)	7 (100%)	

Table 4. The relation between MS types with EDSS patients who their serum uric acid below 3 mg/dl.

MS type with no. of patients uric acid below 3 mg/dl	No. of patients with EDSS less than 3.5	No. of patients with EDSS equal or more than 3.5	
RRMS (18)	4 (22.2%)	14 (77.8%)	
SPMS (7)	0	7 (100%)	

Table 5.The mean serum UA concentration waslower in the MS 47 cases (3.77) than in the samegroup (4.22) after one month of follow up, thedifference is statistically significant (P < 0.01).</td>

Uric acid	First visit	After one month
Mean	3.77	4.22
S.D.	0.98	0.77
95% C.I.	3.48 - 4.06	3.99 - 4.44

Table 6. Correlation of the UA of 47 patients of the cases after 1 month of follow up with different variables.

Correlation between	R-value	<i>p</i> -value
UA and age in years	-0.239	0.106*
UA and duration of illness	-0.197	0.184*
UA and no. of relapses	-0.288	0.05
UA and EDSS score	-0.205	0.168*
UA and clinical activity	0.091	0.542*
UA and MS type	-0.186	0.21*
UA before and after 1 month	0.543	0.01

* Not statistically significant

and clinical measures of MS was not tested in these studies. Constantinescu et al.²⁷ reported elevation in the mean serum UA levels in MS patients after 6 months of treatment with glatiramer acetate, suggesting that beneficial effect of this drug is based on elevation in UA,27 as a natural scavenger of peroxynitrite. We found significantly lower values of serum UA in SPMS patients than in controls, while significant differences were observed neither between RR or PPMS and controls nor between the different clinical subtypes of MS. However, after performing multivariate linear regression analyses, we concluded that low values of UA in SPMS were not due to the independent effect of SP disease course. Our RRMS and SPMS patients may have had lower serum UA concentrations due to the relative predominance of female gender, active disease, and longer disease duration in this subgroup of patients since we found a strong inverse correlation between these variables and serum UA concentration. Inverse correlation between serum UA concentration and female gender may be one of the factors that influence female predominance in MS. As noted above, we found significantly lower concentrations of UA in sera from patients with a clinically active disease than in those with clinically inactive disease or controls. Moreover, we found an independent effect of disease activity on serum UA in MS patients by the multivariate regression analyses. Another study depend on MRI which resulted lower values were also observed in patients with Gd-DTPA enhancement on brain MRI as a sign of MRI disease activity in comparison with those without active lesions, but the difference did not reach statistical significance, perhaps due to the small number of patients in whom brain MRI with Gd-DTPA injection was performed. Since Gd-DTPA enhancement in MRI of the brain in MS is associated with inflammation.²⁸ Whether the reduction in UA level in patients with active MS is a cause or a consequence of disease activity in MS remains uncertain. It may be speculated that patients with active MS have an intrinsically reduced antioxidant reserve which contributes to the development of CNS inflammation and tissue damage in MS, or that CNS inflammation in the active phase of the disease leads to the consumption of UA as a scavenger. Hooper et al.29 recently reported that UA protects the integrity of the blood-CNS barrier in mice with EAE such that inflammatory cell migration into CNS tissues is reduced. However, the same study showed that exogenously administered UA penetrates the already compromised blood-CNS barrier and

blocks peroxynitrite-mediated tyrosine nitration and apoptotic cell death within the areas of inflammation in the spinal cord in EAE, speaking in favor of the latter notion. Although inflammation and demyelination are central features in MS, recent observations from pathological studies and magnetic resonance spectroscopy have led to the hypothesis that axonal damage is responsible for a significant proportion of the clinical phenomena and irreversible neurological impairment in this disease.^{30,31} Axonal damage, represented by decreases in brain N-acetylaspartate concentrations in magnetic resonance spectroscopy studies, was shown to progress over time³² and to be correlated with clinical disability.^{33,34} Serial observations^{10,19,24} indicated that low UA levels are possibly a primary feature of MS patients. Instead, another study suggested that UA levels serve as a marker of MS activity, being inversely correlated with disease activity and duration.²⁵ In the present study, we obtained confirmatory evidence for an association of low serum UA levels with MS, even though some control neurological diseases (such as epilepsy and stroke) were potentially related to hyperuricemia. Drulovic and colleagues²⁵ found confirmatory evidence that patients with active RR and SP forms of MS have significantly lower UA levels than controls. On the other hand, in the overall group (including MS patients with clinically inactive forms of MS) serum UA levels did not differ significantly from those of controls, although a strong tendency was evident (p = 0.068). It may be observed that the two study populations (MS and OND) of Drulovic et al.²⁵ differed in terms of numerosity (240 and 104, respectively) and that about 30% of their control patients had seizures. Since carbamazepine and valproic acid can decrease serum UA levels,35 they may possibly have lowered the mean UA level in the OND population. In their study, comparison of UA levels in patients stratified according to MS course and disability showed a significant difference. This observation may favor the view that, although a concomitant disease activity-related effect is present, low UA levels might represent a primary "MS-specific" deficiency. In fact, despite the fact that the UA level was lower in clinically documented active patients, therefore partially confirming Drulovic et al's²⁵ results, a general mean UA decrease is evident also in clinically and MRI inactive MS patients as compared to OND. Therefore, the question whether reduced UA level in MS is a primary deficit or an epiphenomenon related to its oxidation by PN and free radicals remains open, although the two alternative hypotheses are not mutually exclusive and concomitance of either facts is likely. In the latest metabolic stage, UA is transformed into allantoin which does not have PN scavenging activity. In other pathological conditions, such as myocardial infarction³⁶ and chronic lung disease of preterm infants³⁷ or during muscle metabolic stress induced by intense exercise,³⁸ serum allantoin is a useful early predictor of the subsequent free radicals generation. To our knowledge, no study has assessed allantoin levels in MS patients so as to determine whether UA is primarily deficient or secondarily reduced by virtue of its protective role against oxidant compounds. Investigations aimed at determining such an in vivo surrogate marker of free radical production in MS are clinically relevant.

Conflict of Interest

None

References

- Joseph BG, John B, Nancy JH and June H. An overview of multiple sclerosis. *Multiple Sclerosis*: A Self-Care Guide to Wellness, 2nd Edition, New York, Demos Medical Publishing. 2005; 1-932603-07-7.
- "Uric Acid." Biological Magnetic Resonance Data Bank. Indicator Information (gen_metab_summary_5.php-molName=uric_acid.htm#INCHI) Retrieved on 18 February 2008.
- Lin RF, Lin TS, Tilton RG and Cross AH. Nitric oxide localized to spinal cords of mice with experimental allergic encephalomyelitis: An electron paramagnetic resonance study. J Exp Med. 1993 Aug 1;178(2):643–648.
- Hooper DC, Ohnishi ST, Kean Ŕ, Numagami Y, Dietzschold B and Koprowski H. Local nitric oxide production in viral and autoimmune diseases of the central nervous system. Proc Natl Acad Sci U S A. 1995Jun 6;92(12):5312– 5316.
- Akaike T, Noguchi Y and Ijiri S, et al. Pathogenesis of influenza virus-induced pneumonia: involvement of both nitric oxide and oxygen radicals. Proc Natl Acad Sci U S A. 1996 Mar 19; 93(6):2448–2453.
- Radi R, Beckman JS, Bush KM and Freeman BA. Peroxynitrite-induced membrane lipid peroxidation: the cytotoxic potential of superoxide and nitric oxide. Arch Biochem Biophys. 1991 Aug 1;288(2):481–487.
- Beckmann JS, Ye YZ and Anderson PG, et al. Extensive nitration of protein tyrosines in human atherosclerosis detected by immunohistochemistry. Biol Chem Hoppe Seyler. 1994 Feb;375(2):81–88.
- Ischiropoulos H, Zhu L and Chen J, et al. Peroxynitrite-mediated tyrosine nitration catalyzed by superoxide dismutase. Arch Biochem Biophys. 1992 Nov 1;298(2):431–437.
- Whiteman M and Halliwell B. Protection against peroxynitrite-dependent tyrosine nitration and alpha 1-antiproteinase inactivation by ascorbic acid. A comparison with other biological antioxidants. Free Radic Res. 1996 Sep;25(3):275–283.
- Hooper DC, Bagasra O and Marini JC, et al. Prevention of experimental allergic encephalomyelitis by targeting nitric oxide and peroxynitrite: Implications for the treatment of multiple sclerosis. Proc Natl Acad Sci U S A. 1997 Mar 18;94(6):2528–2533.
- 11. Huang S, Hendriks W and Althage A, et al. Immune response in mice that lack the interferon-gamma receptor. Science. 1993 Mar 19;259(5102):1742–1745.
- 12. Beckman JS. The double-edged role of nitric oxide in brain function and superoxide-mediated injury. J Dev Physiol. 1991 Jan;15(1):53–59.
- Cross AH, Misko TP, Lin RF, Hickey WF, Trotter JL and Tilton RG. Aminoguanidine, an inhibitor of inducible nitric oxide synthase, ameliorates experimental autoimmune encephalomyelitis in SJL mice. J Clin Invest. 1994 Jun;93(6):2684–2690.
- Mikkelsen WM, Dodge HJ and Valkengurg H. The distribution of serum uric acid values in a population unselected as to gout or hyperuricemia: Tecumseh, Michigan 1959-1960. Am J Med. 1965 Aug;39:242–251.
- Smith KJ, Kapoor R and Felts PA. Demyelination: The role of reactive oxygen and nitrogen species. Brain Pathol. 1999 Jan;9(1):69–92.
- Merrill JE, Ignarro LJ, Sherman MP, Melinek J and Lane TE. Microglial cell cytotoxicity of oligodendrocytes is mediated through nitric oxide. J Immunol. 1993 Aug 15;151(4):2132–2141.
- Van der Veen RC and Roberts LJ. Contrasting roles for nitric oxide and peroxynitrite in the peroxidation of myelin lipids. J Neuroimmunol. 1999 Mar 1;95(1-2):1–7.
- Redford EJ, Kapoor R and Smith KJ. Nitric oxide donors reversibly block axonal conduction: Demyelinated axons are especially susceptible. Brain. 1997 Dec; 120 (Pt 12):2149–2157.
- Hooper DC, Spitsin S and Kean RB, et al. Uric acid, a natural scavenger of peroxynitrite, in experimental allergic encephalomyelitis and multiple sclerosis. Proc Natl Acad Sci U S A. 1998 Jan 20; 95(2):675–680.
- Kastenbauer S, Koedel U and Pfister HW. Role of peroxynitrite as a mediator of pathophysiological alterations in experimental pneumococcal meningitis. J Infect Dis. 1999 Oct; 180(4):1164–1170.

- Yu ZF, Bruce-Keller AJ, Goodman Y and Mattson MP. Uric acid protects neurons against excitotoxic and metabolic insults in cell culture, and against focal ischemic brain injury in vivo. J Neurosci Res. 1998 Sep 1; 53(5):613–625.
- 22. Van der Veen RC, Hinton DR, Incardonna F and Hofman FM. Extensive peroxynitrite activity during progressive stages of central nervous system inflammation. *J Neuroimmunol*. 1997 Jul; 77(1):1–7.
- 23. Kean RB, Spitsin SV, Mikheeva T, Scott GS and Hooper DC. Peroxynitrite scavenger uric acid prevents inflammatory cell invasion into the central nervous system in experimental allergic encephalomyelitis through maintenance of blood-central nervous system barrier integrity. J. Immunol. 2000 Dec 1; 165(11):6511–6518.
- 24. Koprowski H, Spitsin SV and Hooper DC. Prospects for the treatment of multiple sclerosis by raising serum levels of uric acid, a scavenger of peroxynitrite. Ann Neurol. 2001 Jan;49(1):139.
- Drulović J, Dujmović I and Stojsavljević N, et al. Uric acid levels in sera from patients with multiple sclerosis. J Neurol. 2001 Feb; 248(2): 121–126.
- Karg E, Klivényi P, Németh I, Bencsik K, Pintér S and Vécsei L. Nonenzymatic antioxidants of blood in multiple sclerosis. J Neurol. 1999 Jul; 246(7):533– 539.
- 27. Constantinescu CS, Freitag P and Kappos L. Increase in serum levels of uric acid, an endogenous antioxidant, under treatment with glatiramer acetate for multiple sclerosis. Mult Scler. 2000 Dec; 6(6):378–381.
- Brück W, Bitsch A, Kolenda H, Brück Y, Stiefel M and Lassmann H. Inflammatory central nervous system demyelination: Correlation of magnetic resonance imaging findings with lesion pathology. *Ann Neurol.* 1997 Nov; 42(5):783–793.
- Hooper DC, Scott GS and Zborek A, et al. Uric acid, a peroxynitrite scavenger, inhibits CNS inflammation, blood-CNS barrier permeability changes, and tissue damage in a mouse model of multiple sclerosis. FASEB J. 2000 Apr;14(5):691–698.
- Coles AJ, Wing MG and Molyneux P, et al. Monoclonal antibody treatment exposes three mechanisms underlying the clinical course of multiple sclerosis. Ann Neurol. 1999 Sep;46(3):296–304.
- Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mörk S and Bö L. Axonal transection in the lesions of multiple sclerosis. N Engl J Med. 1998 Jan 29;338(5):278–285.
- 32. Arnold DL, Riess GT and Matthews PM, et al. Use of proton magnetic resonance spectroscopy for monitoring disease progression in multiple sclerosis. Ann Neurol. 1994 Jul; 36(1):76–82.
- Arnold DL, Matthews PM, Francis G and Antel J. Proton magnetic resonance spectroscopy of human brain in vivo in the evaluation of multiple sclerosis: Assessment of the load of disease. Magn Reson Med. 1990 Apr; 14(1):154– 159.
- Ferguson B, Matyszak MK, Esiri MM and Perry VH. Axonal damage in acute multiple sclerosis lesions. Brain. 1997 Mar; 120 (Pt 3):393–399.
- Ring HA, Heller AJ, Marshall WJ, Johnson AL and Reynolds EH. Plasma uric acid in patients receiving anticonvulsant monotherapy. Epilepsy Res. 1991 Apr; 8(3):241–244.
- 36. Kock R, Delvoux B, Sigmund M and Greiling H. A comparative study of the concentrations of hypoxanthine, xanthine, uric acid and allantoin in the peripheral blood of normals and patients with acute myocardial infarction and other ischemic diseases. Eur J Clin Chem Clin Biochem. 1994 Nov; 32(11):837–842.
- Ogihara T, Kim HS and Hirano K, et al. Oxidation products of uric acid and ascorbic acid in preterm infants with chronic lung disease. Biol Neonate. 1998; 73(1):24–33.
- Hellsten Y, Tullson PC, Richter EA and Bangsbo J. Oxidation of urate in human skeletal muscle during exercise. Free Radic Biol Med. 1997; 22(1-2): 169–174.